Chemometric Tools for NIRS and NIR Hyperspectral Imaging

Laura M. DALE[1] [2], André THEWIS[1], Ioan ROTAR[2], Juan A. FERNANDEZ PIERNA[3], Christelle BOUDRY [1], Roxana M. VIDICAN [2], Vincent BAETEN[3]

[1] Animal Science Unit, Gembloux Agro-Bio Tech, University of Liège, 2, Passage des Déportés, 5030 Gembloux, Belgium; dale_lm@yahoo.com; dale.laura@student.ulg.ac.be; athewis@ulg.ac.be; christelle.boudry@ulg.ac.be.
[2] Department of Grassland and Forage Crops, University of Agricultural Science and Veterinary Medicine Cluj Napoca, 3-5, Calea Manaştur, 400372, Cluj Napoca; rotarioan52@yahoo.fr; roxanavidican@yahoo.com;
[3] Walloon Agricultural Research Centre, Valorisation of Agricultural Products Department, 24 Chaussée de Namur, 5030 Gembloux, Belgium; fernandez@cra.wallonie.be; baeten@cra.wallonie.be.

Abstract. Nowadays in agriculture, new analytical tools based on spectroscopic technologies are developed. Near Infrared Spectroscopy (NIRS) is a well known technology in the agricultural sector allowing the acquisition of chemical information from the samples with a large number of advantages, such as: easy to use tool, fast and simultaneous analysis of several components, non-polluting, non-invasive and non destructive technology, and possibility of online or field implementation. Recently, NIRS system was combined with imaging technologies creating the Near Infrared Hyperspectral Imaging system (NIR-HSI). This technology provides simultaneously spectral and spatial information from an object. The main differences between NIR-HSI and NIRS is that many spectra can be recorded simultaneously from a large area of an object with the former while with NIRS only one spectrum was recorded for analysis on a small area. In this work, both technologies are presented with special focus on the main spectrum and images analysis methods. Several qualitative and quantitative applications of NIRS and NIR-HSI in agricultural products are listed. Developments of NIRS and NIR-HSI will enhance progress in the field of agriculture by providing high quality and safe agricultural products, better plant and grain selection techniques or compound feed industry's productivity among others.

Keywords: NIRS, NIR-HSI, non-destructive methods, chemometric tools, agriculture applications

INTRODUCTION

Near Infrared Spectroscopy (NIRS) is a well known technology in the agricultural sector allowing the acquisition of chemical information from the samples (Rodriguez-Otero, et al., 1997). By NIRS, C-H, N-H and O-H bonds are induced to vibrate. This principle is used to identify and quantify components. NIRS allows the acquisition of the reflectance spectra of opaque milled or intact materials. NIRS is characterized by acquisition of a typical NIR spectrum which can be considered as the spectral signature or spectral fingerprint of the material. However because of NIRS low sensibility different functional groups may be detected in overtones and combinations bands (Burns and Margoshes, 1992). It is an adequate technique for the analysis of major components (chemical composition, microorganism detection and quantification) in agriculture with minimum sample preparation.

NIRS is a well known technology in the agricultural sector since the scientific works regarding soybean moisture of Norris in the 1960's (Hart et al., 1962). In the last years, new areas of work based on the NIRS technology have been developed combining NIR systems

and a microscope to create the NIR microscopy (NIRM) (Baeten et al., 2012). More recently, NIR was combined with imaging technologies creating the Near Infrared Hyperspectral Imaging System (NIR-HSI) (Fernández Pierna et al., 2004; Fernández Pierna et al., 2012). ElMasry and Sun (2010) explained that «hyper» in hyperspectral means «over» or «too many» and it refers to the large number of wavelength bands measured. While, the «spectroscopy» word means «seeing», the «spectrometry» word means «measuring» and the hyperspectral word means «many bands». NIR-HSI system provides spectral and spatial informations from an object that forms a three dimensional "*hypercube*",which can be used to extract physical and chemical information from an object. The images provide sufficient information to identify and distinguish each spectrum as a unique material. NIR-HSI has different instrumentation approaches: point (staring) scan, push-broom (line) scan or plane (global) scan (Fernández Pierna et al., 2009).

Fifteen years ago, only experts in remote sensing spectral images had access to hyperspectral systems or software tools to explore such images. In the last decade, hyperspectral image analysis was developed into one of the most powerful and fastest growing technology in remote sensing (Kavitha et al., 2012).

The objectives of this paper are to describe the advantages and disadvantages of NIRS and NIR-HSI and to display how a spectrum or an image can be analysed. Finally, some applications made by the NIRS and NIR-HSI in agriculture are listed.

ADVANTAGES AND DISADVANTAGES

Major advantages of spectroscopic techniques are: the ease of use, repeatability and reproducibility, reasonable start-up cost, non-polluting, non-invasive and non-destructive analyses and the possibility of online or directly in the field implementation. By NIR-HSI many spectra can be recorded simultaneously for one sample instead of a unique, average spectrum obtained when using classical NIRS (Fig. 1).
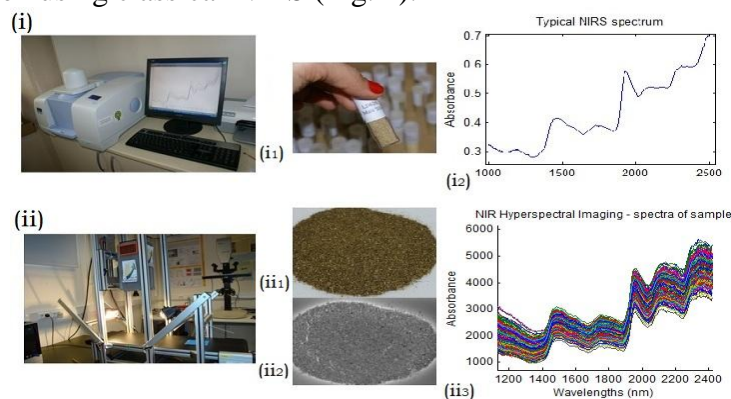

Fig.1. Acquisition of spectrum/spectra by (i) conventional NIRS system and (ii) Laboratory scale NIR-HSI system (source CRA-W, Gembloux, Belgium)

Legend: (i) NIRS system (Forage Crops Lab, USAMV Cluj, Romania); (i$_1$) Sample support for NIRS system; (i$_2$) Typical spectrum of NIRS system; (ii) Laboratory scale NIR-HSI system (source CRA-W, Gembloux, Belgium); (ii$_1$) Photography of sample; (ii$_2$) Hyperspectral image of sample; (ii$_3$) Typical spectra of Laboratory scale NIR-HSI system

Hyperspectral images provide more information i. e. spectral and spatial information, than could be obtained when NIRS technology was used. One spectral image regroups numerous spectra. So each pixel of spectral image corresponds to one spectrum of the target sample. The spectrum or spectral signature can be used to characterize, classify or even identify any given material (Shaw and Manolakis, 2002).

The disadvantages of NIRS and NIR-HSI are: the relatively high price of instruments; the requirement of huge hardware speed; the necessity of calibration models for standardization; the possible presence of pixels (spectrum) in the image which do not contain any chemical information like for example bad, dead, noise, blinking or drifting pixels (Chang, 2000; ElMasry and Sun, 2010).

SPECTRA ANALYSES

Qualitative and quantitative analyses by NIRS usually require the application of calibration algorithms based on physico-chemical measurements. In order to get efficient qualitative and quantitative information from data coming from NIRS and NIR-HSI instrumentation, chemometric tools are necessary (Roggo et al., 2005). Chemometrics is the science of extracting relevant information from measurements made in chemical systems, using mathematical and statistical procedures (Massart et al., 1988).

The building of calibration models starts with spectra pre-processing treatment: after collection of spectra, it is necessary to perform a pre-treatment to remove high- or low-frequency interferences. Different types of pre-processing treatments are: polynomial baseline correction, Savitzky - Golay derivative, Standard Normal Variate (SNV), mean-centering and unit variance normalization among others (Gowen et al., 2007; ElMasry and Sun, 2010).

The most frequent pre-processing treatment used in practical are:
- SNV transformation removes the slope variation from spectra caused by scatter and variation of particle size (Candolfi et al., 1999).
- Derivative conversion, unimportant baseline signal from samples are removed by taking the derivative of the measured responses with respect to the variable number (index) or other relevant axis scale (wavelength, wavenumbers, etc.) (Wise et al., 2006). First derivative is usually used to remove any offset from the sample and de-emphasizing lower-frequency signals (Wise et al., 2006) while second derivative will accentuate the higher-frequency to enhance selectivity (Wise et al., 2006).

After spectra pre-processing treatment the calibration algorithms can be applied for classification and quantification. In order to analyse data, many multivariate analytical tools are used, such as: principal component analysis (PCA), principal component regression (PCR), multi-linear regression (MLR), partial least squares regression (PLS), modified partial least squares regression (MPLS), partial least squares discriminant analysis (PLS-DA), linear discriminant analysis (LDA), support vector machines (SVM), artificial neural networks (ANN), soft independent modeling of class analogy (SIMCA) baseline shift (BLS), spectral information and divergence (SID) (Chang, 2000; Fernandez Pierna et al., 2006; Gowen et al., 2007; Gómez - Sanchis et al., 2012).

The most frequent mathematical algorithms used in practice are:
- PCA: used for data compression and information extraction (Wise et al., 2006);
- MLR: allowed to establish a link between a reduced number of wavelengths (or wavenumber) and a property of the samples (Roggo et al., 2005) and to find a single factor that best correlates predictor (X) variables with predicted (Y) ones (Wise et al., 2006);
- PLS: used to establish a linear link between two matrices, the spectral data X and the reference values Y (Roggo et al., 2005); in other words, it attempted to find factors for both capture variance and also to achieve correlation (Wise et al., 2006), while PLS-DA is performed to discriminate between classes (McGoverin et al., 2011);
- ANN: it can be applied for pattern recognition, classification or clustering and quantitative modelling (Dolmatova et al., 1997);
- SVM: used for solving problems of nonlinear classification/regression, model estimation and density estimation (Fernandez Pierna et al., 2006).

The spectral pre-processing treatments and calibration algorithms are extensively reviewed in the literature (Dolmatova et al., 1997; Roggo et al., 2005; Fernandez Pierna et al., 2006; Wise et al., 2006; ElMasry and Sun, 2010; Yao and Lewis, 2010; Fernandez Pierna et al., 2011).

Assessment of calibration performance: NIR calibration models performances can be characterized by several parameters: standard error of calibration (SEC) or standard error of cross validation (SECV). To perform calibration model performance, an independent set of samples is used to get the standard error of prediction (SEP) and the squared coefficient of correlation (RSQ), which are used to describe the NIR analytical error when analyzing samples of unknown quantitative composition (Hartmann and Buning-Pfaue, 1998).

APLICATIONS IN AGRICULTURE OF NIRS AND NIR-HSI

The first studies based on the NIRS technique were published between 1930 and 1940. By 1990, more than 1000 articles were published using this technique (Burns and Margoshes, 1992). If we look in the agriculture applications more than 1000 are carried out in 2012 (Google Scholar, consulted on 10.05.2012).

Tab. 1.

Summary of measurement mode, product type, wavelength region used and classification algorithm employed in papers published on NIRS and NIR-HSI applications in agriculture and agro-industries

NIRS system applications				NIR Hyperspectral Imaging applications			
Products	Wavelength (nm)	Model	Reference	Products	Wavelength (nm)	Model	Reference
Forage quality : - ash content - digestibility - protein content	1100-2500	MLR MLR PLS	Vazquez de Aldana et al., 1996; Vidican et al., 2000; Hărmanescu and Moisuc, 2009	Forage quality : - protein content - digestibility	1100-2500	PLSDA	Dale et al., 2012
Compound feedstuffs	400-2500	MPLS	Pérez-Marin et al., 2004	Compound feed and impurities	1100-2500	PCA SVM	Fernandez Pierna et al., 2006 and 2012
Wheat quality - protein content	1100-2500	PCA SIMCA	Gatius et al., 2004	Barley, wheat and sorghum grains - moisture content	1000-2500	PCA PLSDA	McGoverin et al., 2011
Cereals: - modifications made by filamentous fungi and yeasts - damages made by *Fusarium culmorum*	1000-2500 570-1100	Review PCA	Santos et al., 2010 Pettersson and Aberg, 2003	Detection of Fusarium - in maize - in wheat	720-940 400-1000	ANN PCA	Firrao et al., 2010; Bauriegel et al., 2011
Detection of ergot bodies in wheat kernels	1100-2400	Fisher coef. value	Vermeulen et al., 2009	Detection of ergot bodies in wheat kernels	900-1700	Fisher coef. value	Vermeulen et al., 2009
Hardness in diverse corn germplasm	1000-2500	BLS	Hoffman et al., 2010	Glassy and floury endosperm in maize	1000-2498	PLSDA PCA	Williams et al., 2009
Alphamylase activity - in malted barley - in wheat kernel	400-2500 1235-2450	PLS MLR PCA PLS	Tarr et al., 2012 Xing et al., 2011	Alphamylase activity - in wheat kernel	1235-2450	PCA PLS	Xing et al., 2011
Potato constituents	1100-2500	PCA MPLS	Hartmann and Buning-Pfaue, 1998	Detection of hollow heart in potatoes	900-1700	SVM	Dacal-Nieto et al., 2011
Detection of castor bean meal in flour-containing products	1000-2500	MPLS	Rodriguez-Saona et al., 2000	Detection of meat and bone meal in compound feeds	960-1662	SVM	Fernandez Pierna et al., 2004
Detection and identification of bacteria in an isolated system	750–2500	PCA PLSDA SIMCA	Alexandrakis et al., 2008	Beet cyst nematodes Microorganism in Spinach (*E. coli*)	1100-2400 400-1000	SVM PCA ANN	Vermeulen et al., 2011; Fernandez Pierna et al., 2012 Siripatrawan et al., 2011

The first use of NIR in agriculture was for the determination of moisture in soybean (Norris et al., 1976), in the case of HSI system remote sensing the first study concerned the detection and mapping of vegetation and minerals and for NIR - HSI the first case study was the detection of meat and bone meal in compound feeds (Fernandez Pierna et al., 2004). Nowadays NIRS and NIR-HSI are used in all agricultural and agro-industries domains from a large scale to a microscopic level. In table 1, some applications that were carried out in agriculture with NIRS and NIR-HSI systems, are enumerated, in order to illustrate the diversified board area of applications: animal nutrition, plant protection, food and feed quality and safety.

CONCLUSIONS

This short review intents to highlight that NIR spectroscopy and the most recent NIR-HSI systems are extremely reliable, non-destructive and rapid techniques for the prediction of quantitative and qualitative chemical and physical properties. However these techniques need the use of Chemometric tools, preprocessing treatment and assessment of calibration models in order to extract the maximum of information these techniques can provide.

Low-cost NIR-HSI systems will be necessary for future developments. Such as NIR-HSI that could easily identify optimal wavelengths/wavebands according to the different applications. For this reason more robust calibration and validation models are necessary in order to find a model that adequately represents the data. Reliability of the models will encourage more widespread online or on-field utilization of this technology (miniaturization or portable instruments) in agriculture and hence, could improve agricultural productivity and reduce the cost of process monitoring and product inspection.

REFERENCES

1. Alexandrakis, D., Downey, G., and Scannell, A. G. M. 2008. Detection and identification of bacteria in an isolated system with near-infrared spectroscopy and multivariate analysis. *J Agric Food Chem*, 56, 3431−3437.
2. Baeten, V., Fernandez Pierna, J.A., Michotte Renier, A., and Dardenne, P. (2005). Imagerie proche infrarouge: analyse de l'alimentation animale, *Techniques del'Ingénieur* (3) RE 34-1-8.
3. Bauriegel, E., Giebel, A., Geyer, M., Schmidt, U., and Herppich, W.B. (2011) Early detection of *Fusarium* infection in wheat using hyper-spectral imaging. *Comput Electron Agr*, 75: 304-312.
4. Burns, D. A., and Margoshes, M. (1992). Historical development. In Burns, D. A., and Ciurczak, E. W., Ed(s): *Handbook of near infrared analysis*, Chapter 1, Marcel Dekker Inc.: New York.
5. Candolfi, A., De Maesschalck, R., Jouan-Rimbaud, D., Hailey, P. A., and Massart, D. L. (1999). The influence of data pre-processing in the pattern recognition of excipients nearinfrared spectra, *J Pharmaceut Biomed*, 21: 115-132.
6. Chang, C. I. (2000). An information theoretic - based approach to spectral variability, similarity and discriminability for hyperspectral image analysis. *IEEE Trans on Inf Theory*, 46(5): 1927-1932.
7. Dacal-Nieto, A., Formella, A., Carrión, P., Vazquez-Fernandez, E., and Fernández-Delgado, M. 2011. Non-destructive detection of hollow heart in potatoes using Hyperspectral Imaging. In Berciano, A., et al., Ed(s): *CAIP'11*, Springer-Verlag, Berlin, Heidelberg, 180-187.
8. Dale, L. M., Fernández Pierna, J. A., Vermeulen, P., Lecler, B., Bogdan, A., Pacurar, F., Rotar, I., Théwis, A., and Baeten, V. (2012). Research on crude protein and digestibility content of *Arnica montana L.* using conventional NIR spectrometry and hyperspectral imaging NIR. *JFAE*,10(1):392-396.
9. Dolmotova, L., Ruckebusch, C., Dupuy, N., Huvenne, J. P., and Legrand, P. (1997). Quantitative analysis of paper coatings using artificial neural network. *Chemometr Intell Lab*, 36: 125-140.

10. ElMasry, G., and Sun, D.-W. (2010). Principles of Hyperspectral Imaging Technology. In: Sun, D. Ed(s): *Hyperspectral Imaging for Food Quality Analysis and Control*, Academic Press, San Diego:3-43.

11. Fernández Pierna, J. A., Baeten, V., and Dardenne, P. (2006). Screening of compound forages using NIR Hyperspectral data. *Chemometr Intell Lab*, 84: 114-118.

12. Fernández Pierna, J. A., Michotte Renier, A., Baeten, V., and Dardenne, P. (2004). IR Camera and Chemometrics (SVM): the winner combination for the detection of MBM. *Stratfeed Symp*, Namur:39.

13. Fernandez Pierna, J.A., Baeten, V., Dardenne, P., Dubois, J., Lewis, E. N., and Burger, J. (2009). Spectroscopic Imaging. In: Brown, S. D., Tauler, R., Walczak, B. Ed(s): *Comprehensive chemometrics. Chemical and Biochemical Data Analysis*, Elsevier, Slovenia: 173-192.

14. Fernandez Pierna, J.A., Vermeulen, P., Amand, O., Tossens, A., Dardenne, P., and Baeten, V. (2012). NIR Hyperspectral Imaging spectroscopy and chemometrics for the detection of undesirable substances in food and feed. *Chemometr Intell Lab*, doi: 10.1016/j.chemolab.2012.02.004.

15. Firrao, G., Torelli, E., Gobbi, E., Raranciuc, S., Bianchi, G., and Locci, R., (2010) Prediction of milled maize fumonisin contamination by multispectral image analysis. *J of Cereal Sci*, 52: 327-330.

16. Gatius, F., Lloveras, J., Ferran J., and Puy, J. (2004). Prediction of crude protein and classification of the growth stage of wheat plant samples from NIR spectra. *The J of Agricultural Sci*, 142(5): 517-524.

17. Gómez-Sanchis, J., Martín-Guerrero, J. D., Soria-Olivas, E., Martínez-Sober, M., Magdalena-Benedito, R., and Blasco, J. (2012). Detecting rottenness caused by Penicillium genus fungi in citrus fruits using machine learning techniques. *Expert Syst Appl*, 39: 780-785.

18. Gowen, A. A., O'Donnell, C. P., Cullen, P. J., Downey, G., and Frias, J. M. (2007). Hyperspectral Imaging - an emerging process analytical tool for food quality and safety control. *Trends Food Sci Tech*, 18: 590-598.

19. Hart, J., Norris, K., and Golumbie, C. (1962). Determination of the moisture content of seeds by near-infrared spectroscopy, *Cereal Chem*, 39: 94-99.

20. Hartmann, R. and Buning-Pfaue, H. (1998). NIR determination of potato constituents. *Am J Potato Res*, 41: 327-334.

21. Hărmănescu, M., and Moisuc, A. (2009). Modelul PLS-CV pentru determinarea conţinutului total de proteină brută din furajele provenite de pe o pajişte permanentă (Grădinari; Caraş - Severin) utilizând spectroscopia NIR (1100-2200 nm). *RJAS*, 41 (1): 161-165.

22. Hoffman, P. C., Ngonyamo-Majee, D., and Shaver, R. D. (2010). Technical note: Determination of corn hardness in diverse corn germplasm using near-infrared reflectance baseline shift as a measure of grinding resistance. *J Dairy Sci*, 93:1685-1689.

23. http://scholar.google.com - consulted on 10.05.2012

24. Kavitha, K., Arivazhagan, S., and Dhivya Priya, R. (2012). Hyperspectral image classification using M-band wavelet transform. *IJAET*, 3(1): 525-533.

25. Massarat, D. L., Vandeginste, B. G. M., Buydens, L. M. C., De Jong, S., Lewi, J. P., Smeyers-Verbeke, J. (1988). Chemometrics: A Textbook; Elsevier: Amsterdam, vol. 2.

26. McGoverin, C. M., Engelbrecht, P., Geladi, P., and Manley M. (2011). Characterisation of non-viable whole barley, wheat and sorghum grains using near-infrared hyperspectral data and chemometrics. *Anal Bioanal Chem*, 401:2283-2289.

27. Norris, K., Barnes, R., Moore, J., and Shenk, J. (1976). Predicting forage quality by infrared reflectance spectroscopy. *J Anim Sci*, 43, 889 -897.

28. Pérez-Marin, D. C., Garrido-Varo, A., Guerrero-Ginel, J.E., Gómez-Cabrera, A. (2004). Near-infrared reflectance spectroscopy (NIRS) for the mandatory labelling of compound feedingstuffs: chemical composition and open-declaration. *Anim Feed Sci Tech*, 116: 333-349.

29. Pettersson, H., and Aberg, L. (2003). Near infrared spectroscopy for determination of mycotoxins in cereals. *Food Control*, 14: 229-232.

30. Rodriguez-Otero, J.L., Hermida, M., and Centeno, J. (1997). Analysis of dairy products by near-infrared spectroscopy: a review. *J Agric Food Chem*, 45: 2815-2819.

31. Rodriguez-Saona, L. E., Fry, F. S., and Calvey, E. M. (2000). Use of Fourier Transform Near-Infrared Reflectance Spectroscopy for Rapid Quantification of Castor Bean Meal in a Selection of Flour-Based Products. *J Agric Food Chem*, 48: 5169-5177.
32. Roggo, Y., Edmond, A., Chalus, P., and Ulmschneider, M. (2005). Infrared Hyperspectral Imaging for qualitative analysis of pharmaceutical solid forms. *Anal Chim Acta*, 535: 79-87.
33. Santos, C., Fraga, M. E., Kozakiewicz, Z., and Lima, N. (2010). Fourier transform infrared as a powerful technique for the identification and characterization of filamentous fungi and yeasts. *Res Microbiol*, 161: 168-175.
34. Shaw, G. and Manolakis, D. (2002). Signal processing for hyperspectral image exploitation. *IEEE Signal Process Mag*, 19(1): 12-16.
35. Siripatrawan, U., Makino, Y., Kawagoe, Y., and Oshita, S. (2011) Rapid detection of *Escherichia coli* contamination in packaged fresh spinach using Hyperspectral Imaging. *Talanta*, 85: 276-281.
36. Tarr, A., Diepeveen, D., and Appels, R. (2012). Spectroscopic and chemical fingerprints in malted barley, *J Cereal Sci*, doi:10.1016/j.jcs.2012.02.007.
37. Vazquez de Aldan, B., Garcia-Criado, B., García-Ciudad, A., and Pérez-Corona, M. E. (1996). Non-destructive method for determining ash content in pasture samples: Application of near infrared reflectance spectroscopy. *Soil Sci and Plant Analy*, 27 (3-40): 795 - 802.
38. Vermeulen, P., Fernández Pierna, J. A., Sinnaeve, G., Dardenne, P. and Baeten, V. (2009) Detection of ergot bodies in cereals by NIRS and Hyperspectral NIR Imaging. Poster in: *14th ICNIRS*, Bangkok, Thailand, 7-13 Nov.
39. Vermeulen, P., Fernández Pierna, J., Tossens, A., Amand, O., Dardenne, P., and Baeten, V. (2011) Identification and quantification of cyst nematode in sugar beet seeds by Hyperspectral NIR Imaging. *14th ICNIRS*, Bangkok, Thailand, 7-13 Nov, 973-977.
40. Vidican, R. M., Rotar, I., and Sima, N. F. (2000) Tehnica NIRS (Near Infrared Reflectance Spectroscopy) şi aplicaţiile sale în analiza calităţii furajelor. *"Agriultura si alimentaţia"*, USAMV Cluj Napoca: 187-191.
41. Williams, P., Geladi, P., Fox, G., and Manley, M. (2009) Maize kernel hardness classification by Near Infrared (NIR) Hyperspectral Imaging and multivariate data analysis. *Analy Chim Acta*, 653: 121-130.
42. Wise, B. M., Shaver, J. M., Gallagher, N. B., Windig, W., Bro, R., and Koch, R. S. (2006). *PLS_Toolbox Version 4.0 for use with Matlab^{TM}*. Wenatchee, WA, USA: Eigenvector Research Inc. 420p.
43. Xing, J., Symons, S., Hatcher, D., and Shahin, M. (2011) Comparison of Short-Wavelength Infrared (SWIR) Hyperspectral Imaging System with an FT-NIR Spectrophotometer for predicting alphaamylase activities in individual Canadian Western Red Spring (CWRS) wheat kernels. *Biosys Eng*, 108: 303-310.
44. Yao, H., and Lewis, D. (2010). Spectral Preprocessing and Calibration Techniques, In: Sun, D.-W. Ed(s): *Hyperspectral Imaging for Food Quality Analysis and Control*, Academic Press, San Diego: 45-78.