

Individual authentication of cocoa beans using Vis-NIR spectroscopy

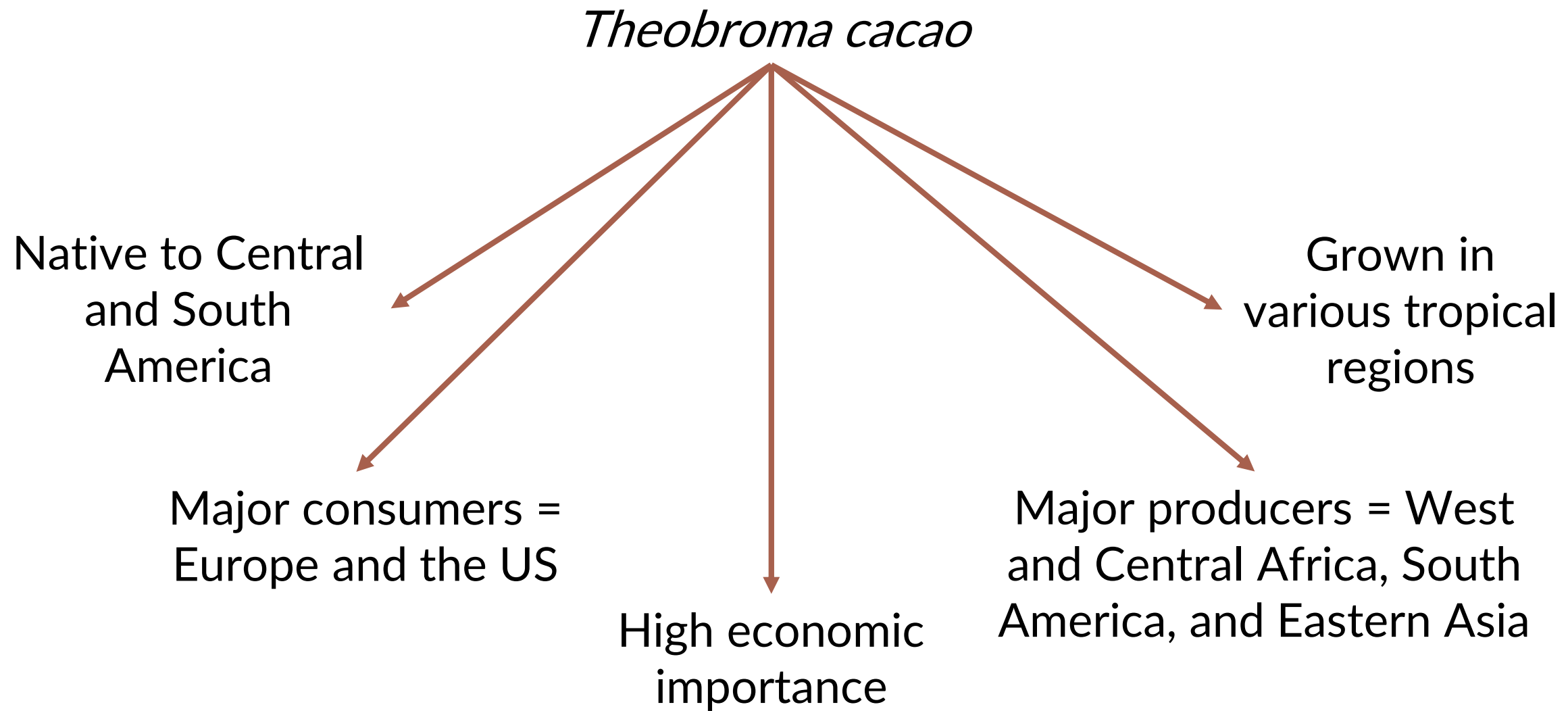
Authors: Antoine Deryck^a, Anne Pinto^b, Giulia Victória Lima^b, Juan Antonio Fernández Pierna^a, Vincent Baeten^a, Hervé Rogez^b

^a Walloon Agricultural Research Center (CRA-W), Knowledge and Valorization of Agricultural Products Department

^b Federal University of Pará (UFPA), Center for Valorization of Amazonian Bioactive Compounds (CVACBA)



Context



Context

Theobroma cacao

Native to Central and South America

Grown in various tropical regions

Major consumers = Europe and the US

High economic importance

Major producers = West and Central Africa, South America, and Eastern Asia

- Brazil = 5th consumer and 6th producer globally
Half from Amazonia region (mainly state of Pará)



Cocoa processing



1. Grow cocoa



2. Harvest cocoa pods



3. Break cocoa pods to extract unfermented beans



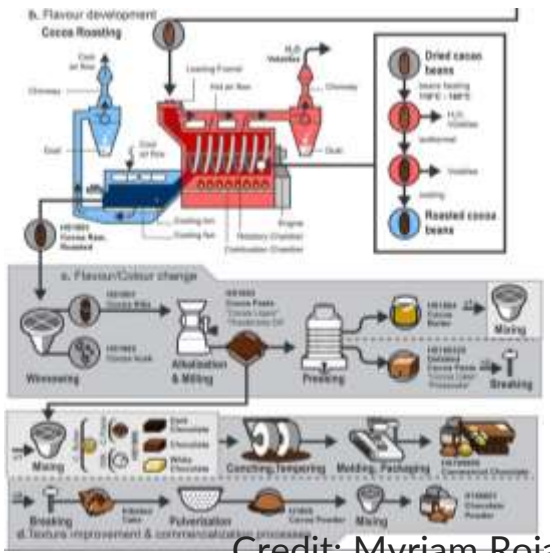
4. Fermentation of beans and mucilage ⚠



5. Drying of beans ⚠



6. Collect fermented and dried beans



7. Industrial steps

Importance of genotypes

- Composition (and later flavor/quality)

Genotype	#BRIX exter	#BRIX inter	pH externo	pH inter	L	a	b	h	Protein full	Lipids Full	Ash
PA169	3.98	13.0	5.83	4.92	26.73	18.21	12.19	33.8835	17.89	36.64	2.03
CCN51	3.43	13.0	5.87	5.08	23.52	15.88	15.85	44.97921	16.62	30.26	2.60
CAB324	3.01	11.0	6.02	5.15	20.34	16.34	14.39	41.46706	16.40	30.50	2.37
P7	2.85	10.0	5.82	5.11	22.04	13.82	14.80	32.68058	18.33	31.61	1.91
CAB499	2.96	11.0	5.98	5.11	26.17	16.66	10.70	32.81032	18.59	32.85	2.21
CA6	3.20	12.0	6.08	5.21	23.90	14.72	17.16	35.21722	18.57	29.74	2.03
PA121	3.66	14.0	5.76	4.94	22.90	15.43	15.28	44.64245	18.09	34.78	1.99
PA195	5.27	11.0	5.65	5.08	19.95	13.94	13.05	43.04929	18.43	30.64	2.01
BE10	3.72	13.5	5.79	4.85	23.41	15.37	11.74	37.5485	16.71	31.67	1.96
CAB270	2.78	15.0	6.13	5.16	25.46	15.31	15.80	41.52663	18.80	29.76	2.20
RB40	5.09	11.0	6.04	5.32	23.98	15.04	15.39	46.37243	20.29	30.37	2.30
RB36	4.35	12.0	5.91	4.87	22.59	15.56	12.78	39.89864	18.44	30.05	1.91
CAB208	4.45	12.5	6.33	5.17	26.01	15.86	17.56	48.11005	17.70	26.31	1.81
IMC67	3.37	14.0	6.15	4.91	22.18	13.81	14.67	47.18841	17.05	31.30	2.50
MO1	3.78	12.0	5.91	5.01	20.64	14.95	14.43	43.93385	16.74	28.34	2.26
CAB214	4.39	9.5	5.93	5.12	17.42	15.40	13.52	41.42286	18.94	29.51	1.77
MA11	4.96	11.5	5.75	4.82	24.44	16.44	13.09	38.86	19.15	35.40	1.98
MA15	3.08	13.0	5.81	4.99	24.41	15.88	12.58	38.76865	18.97	30.27	2.33

- Disease-resistant



- High yield



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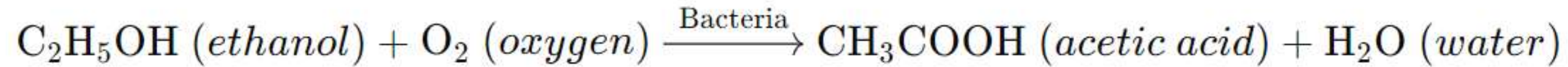
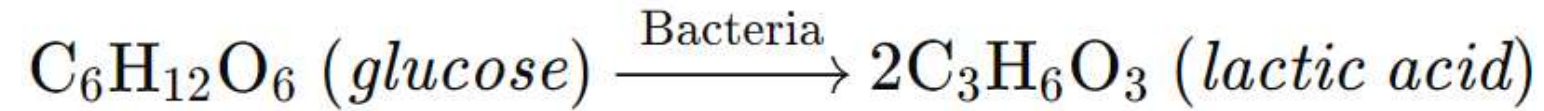
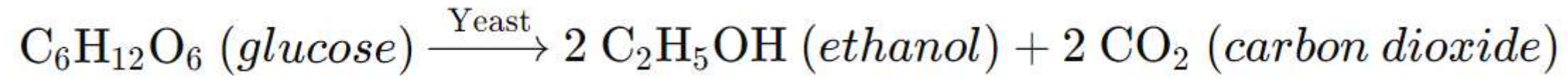
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- High number of genotypes commonly planted in Pará
- Co-planting of genotypes and blending of beans makes it hard to identify high-economic value genotypes
- Beans size but **not accurate**
- Identification by PCR but **costly, destructive, and time-consuming**

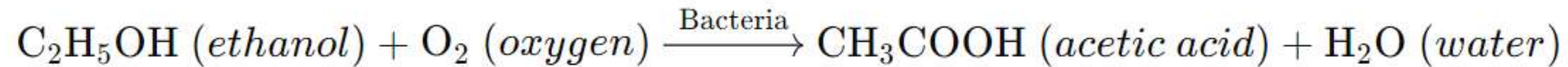
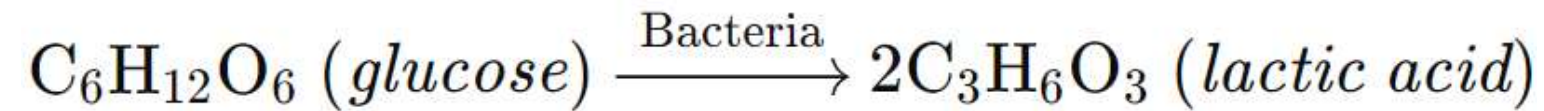
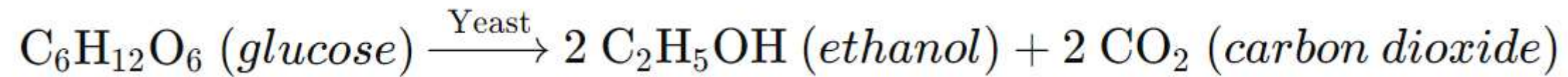
Importance of fermentation

1

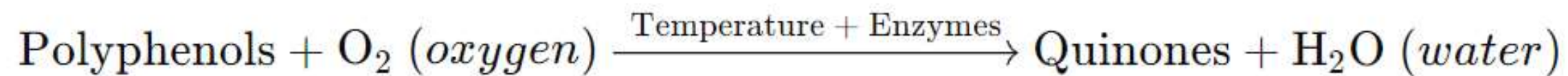
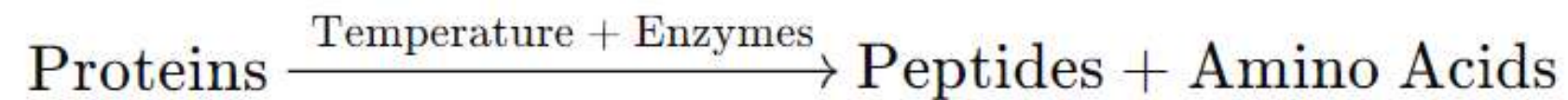


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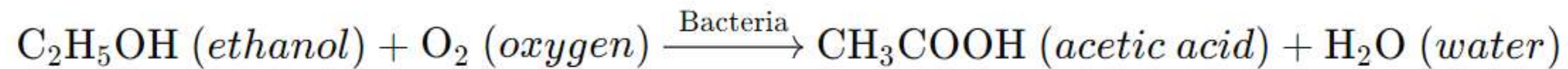
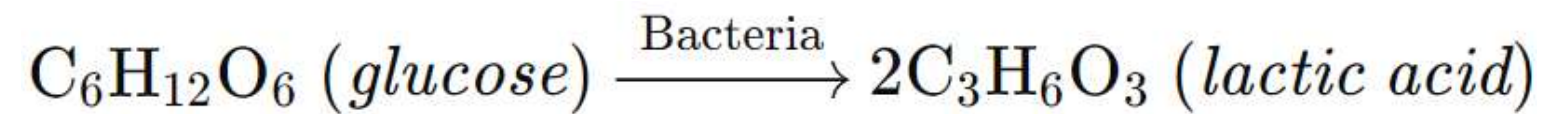
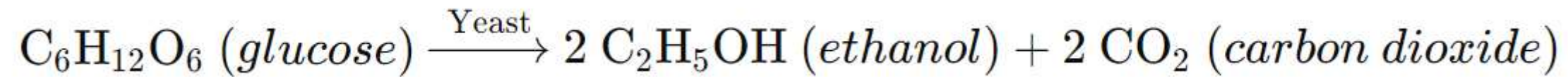


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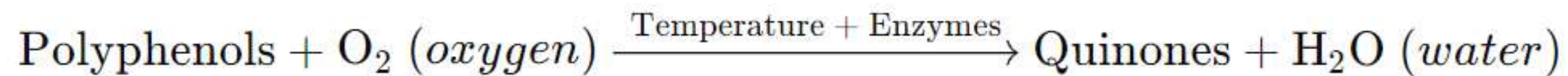
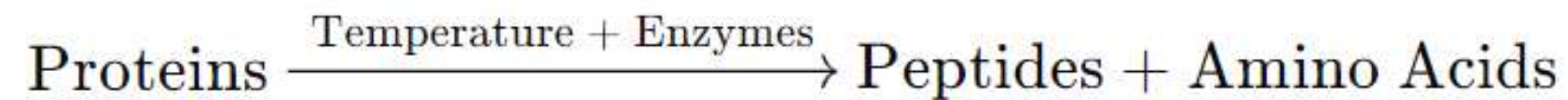


Importance of fermentation

1



2

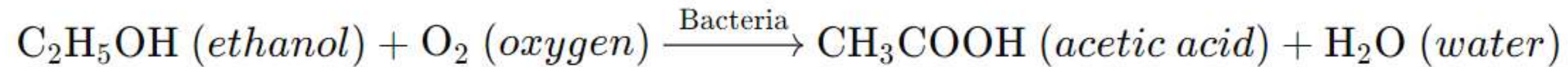
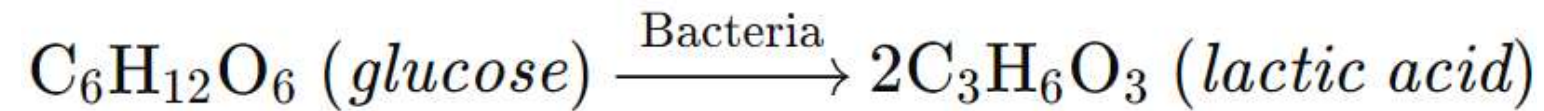
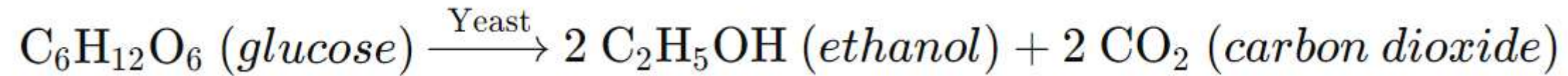


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Flavor
Texture
Composition
Quality

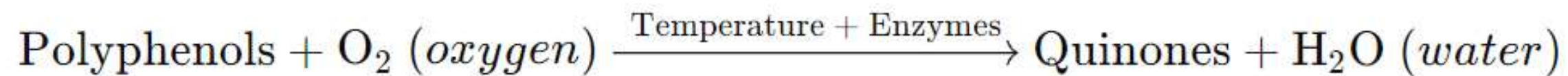
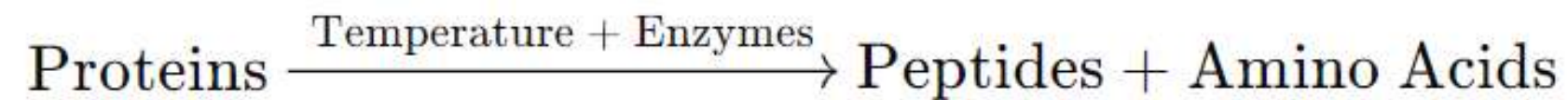


Importance of fermentation

1



2



Color
Flavor
Texture
Composition
Quality



Blending of fermented and unfermented beans → Visual ID by cut-test but **destructive** and **subjective**

Objectives of the study

Efficacy of Vis-NIR spectroscopy for the non-destructive discrimination of

Fermented from unfermented
beans

Forastero cocoa genotypes from
the Eastern Amazonia

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DOI: 10.1007/s12161-017-1137-2
Juliana Hashimoto · Jéssica C. Lima · Renata M. S. Ceieghini · Show all 2 authors · Juliana Pallone



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J.P. Cruz-Tirado^a, Juan Antonio Fernández Pierna^b, Hervé Rogez^c, Douglas Fernandes Barbin^a, Vincent Baeten^b

J Food Sci Technol
https://doi.org/10.1007/s13197-018-3163-5

CrossMark
ORIGINAL ARTICLE

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Douglas Fernandes Barbin¹, Leonardo Fonseca Maciel^{2,3}, Carlos Henrique Vidigal Bazoni³, Margareth da Silva Ribeiro², Rosemary Duarte Sales Carvalho², Eliete da Silva Bispo², Maria da Pureza Spinola Miranda², Elisa Yoko Hirooka³

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————— BUT focus on maximum 5 genotypes

Data collection

Sample collection

Sample preparation

Analyses

- 19 Forastero genotypes from Eastern Amazonia (high importance)

Genotype	Origin	Q1	Genetic group	Q2	Genetic group
CA6	Medicilândia	0.3239	Iquitos	0.2041	Nanay
PA169	Tucumã	0.4665	Marañón	0.2535	Amenolado
PA121	Medicilândia	0.9240	Marañón	0.924	Marañón
PA195	Tucumã	0.6744	Marañón	0.6744	Marañón
BE10	Medicilândia	0.3022	Nanay	0.2576	Manañón
CAB499	Tucumã	0.5731	Purús	0.5731	Purús
CCN51	Medicilândia	0.4608	Criollo	0.2930	Iquitos
IMC67	Medicilândia	0.6602	Iquitos	0.6602	Iquitos
CAB324	Tucumã	0.4620	Purús	0.4598	Nanay
CAB214	Medicilândia	0.5386	Purús	0.4155	Contamana
MA11	Tucumã	0.4192	Purús	0.2835	Amelonado
P7	Medicilândia	0.5295	Nanay	0.4257	Contamana
RB36	Tucumã	0.9511	Purús	0.9511	Purús
RB40	Medicilândia	0.8646	Purús	0.8646	Purús
CAB270	Medicilândia	0.3238	Purús	0.2792	Guiana
MO1	Medicilândia	0.3729	Amelonado	0.2650	Purús
CAB208	Medicilândia	0.7415	Purús	0.7415	Purús
MA15	Medicilândia	0.8389	Purús	0.8389	Purús
CAB314	Tucumã	0.4860	Purús	0.3489	Nanay

- 19x70 fruits collected by CEPLAC in July 2020

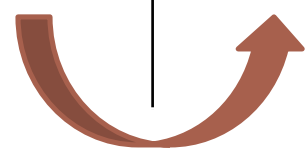
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- 19x70 fruits collected by CEPLAC in July 2020



Sample preparation

- +/- 1 kg beans per genotype
Unfermented



- +/- 1 kg beans per genotype
Fermented (6 days)



- Sun-dried (5 days – moisture < 8%) and stored in refrigerator

Analyses

Data collection

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Sample preparation

- +/- 1 kg beans per genotype Unfermented



- +/- 1 kg beans per genotype Fermented (6 days)



- Sun-dried (5 days - moisture < 8%) and stored in refrigerator

Analyses

- Spectral: 10 random beans per genotype (20 for replicates) with Foss XDS 400-2500 nm (each 2 nm)



- Parameters: Standard analytical methods


Data treatment – Fermentation



Exploratory analyses	Datasets preparation	Models
<ul style="list-style-type: none">• Mahalanobis distance and Z-score method for outliers – none discarded• 4 preprocessing combinations: SNV, SG ($w = 21$, $d = 1$, $p = 2$), SNV + SG and SG + SNV• PCA of raw and preprocessed spectra		

Data treatment – Fermentation

Exploratory analyses	Datasets preparation	Models
<ul style="list-style-type: none">• Mahalanobis distance and Z-score method for outliers – none discarded• 4 preprocessing combinations: SNV, SG ($w = 21, d = 1, p = 2$), SNV + SG and SG + SNV• PCA of raw and preprocessed spectra	<ul style="list-style-type: none">• CreateDataPartition for hold-out validation (proportion of fermentation or not in cal/val)• Calibration dataset = 280 beans (147 F, 133 U)• Validation dataset = 120 beans (63 F, 57 U)	

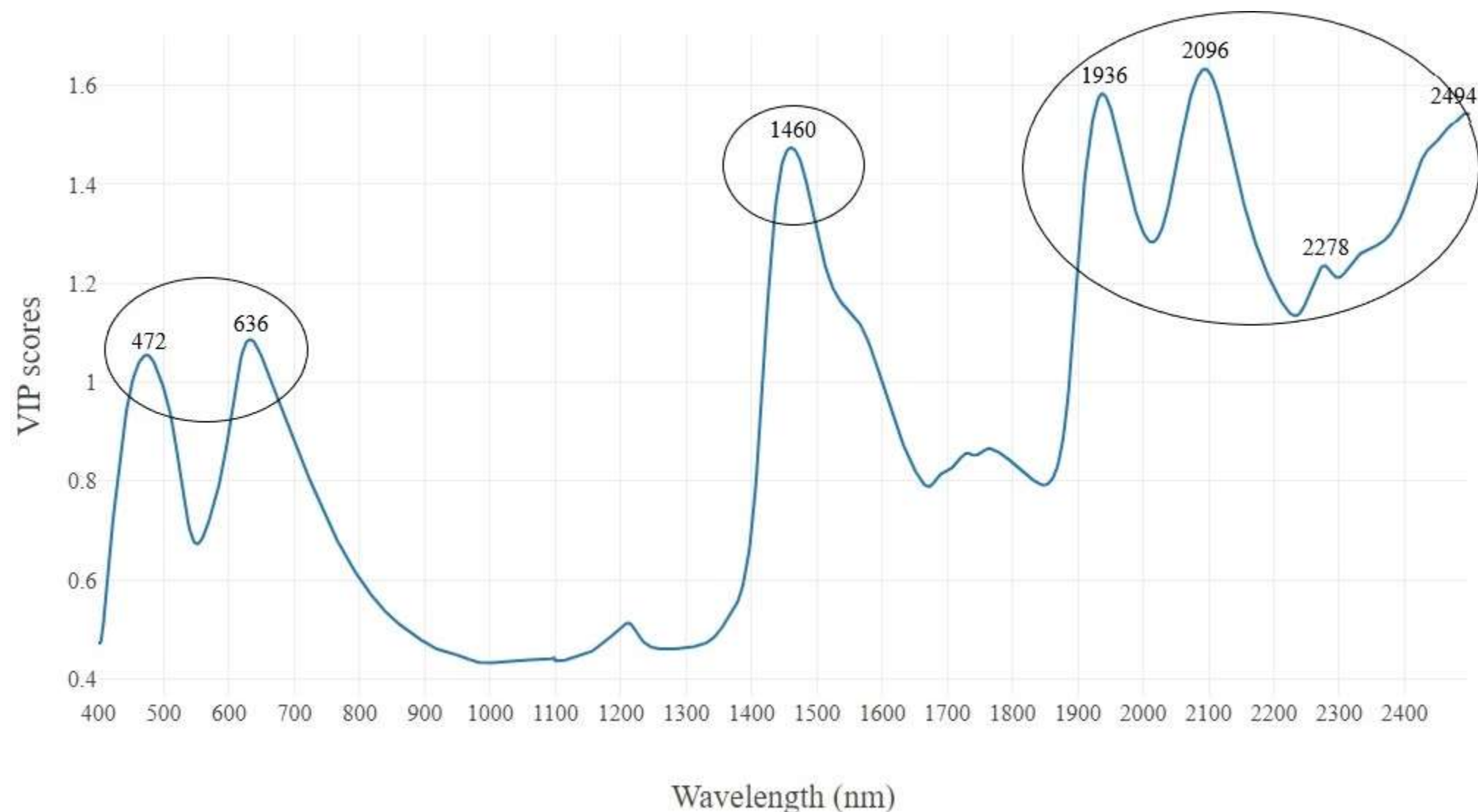


Data treatment – Fermentation



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Results - Fermentation



	PLS-DA						LDA	
	400-2500 nm	400-700 nm	1400-1600 nm	1900-2500 nm	2000-2250 nm	2250-2350 nm	472 + 636 nm	2096 + 2278 nm
NLV	3	3	2	3	2	2	/	/
Sensitivity	1	1	1	1	1	1	0.937	1
Specificity	1	1	0.984	1	0.984	0.984	0.982	1
Accuracy	1	1	0.992	1	0.992	0.992	0.958	1

Data treatment – Genotypes

Exploratory analyses

Datasets preparation

Models

- Mahalanobis distance and Z-score method for outliers – none discarded
- 4 preprocessing combinations: SNV, SG ($w = 21$, $d = 1$, $p = 2$), SNV + SG and SG + SNV
- PCA + t-SNE of raw and preprocessed spectra

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CHEMOMETRICS




RESEARCH ARTICLE

Use of *t*-distributed stochastic neighbour embedding in vibrational spectroscopy

François Stevens ✉, Beatriz Carrasco, Vincent Baeten, Juan A. Fernández Pierna


First published: 23 March 2024 | <https://doi.org/10.1002/cem.3544>

Data treatment - Genotypes

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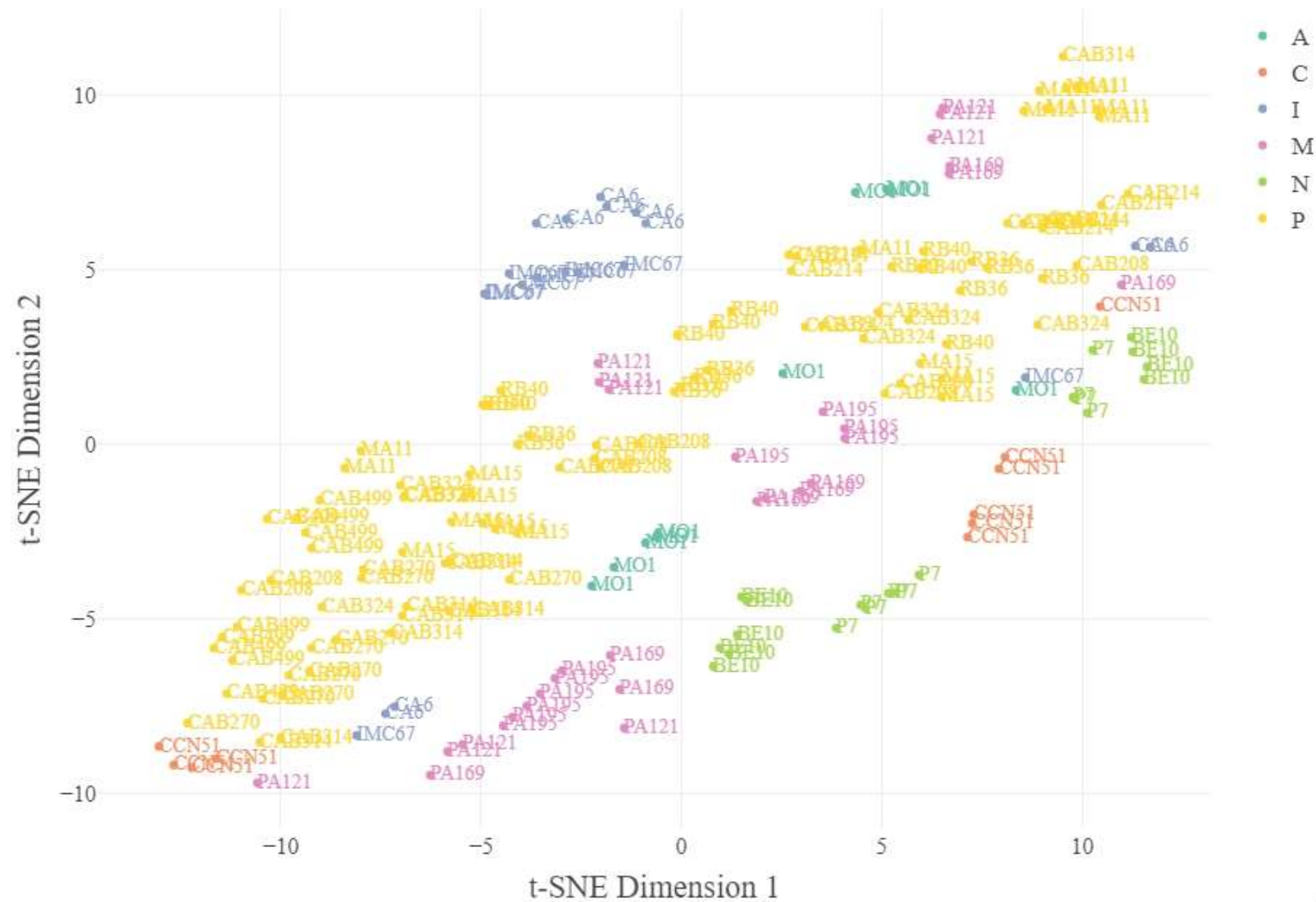


Data treatment - Genotypes

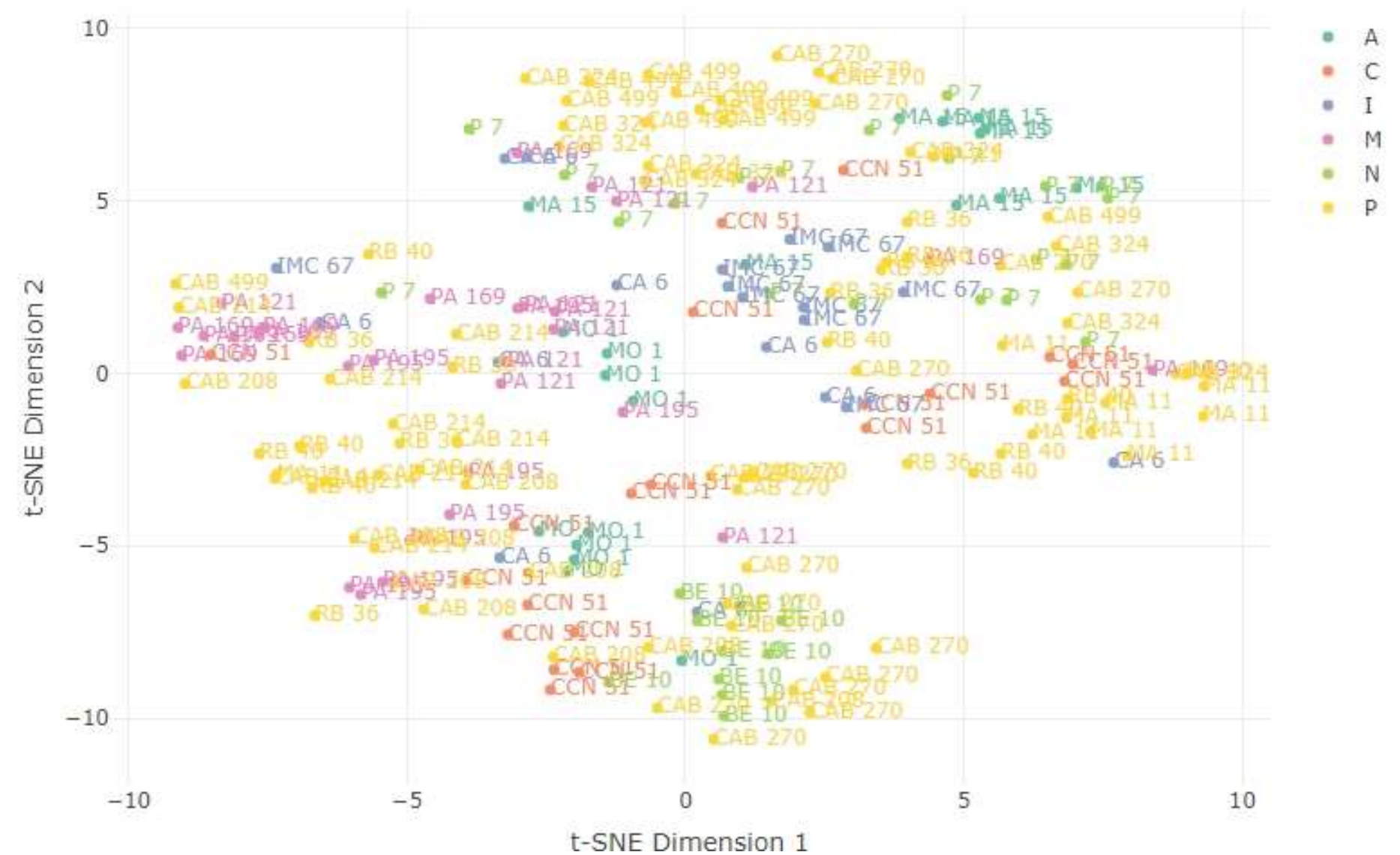
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Results - Genotypes


- Unfermented model: Accuracy = 0,84 (Raw)
9 misclassified \longrightarrow 6 sharing genetic ancestry



- Fermented model: Accuracy = 0,86 (SG)
9 misclassified \longrightarrow 1 sharing genetic ancestry



Conclusion

Fermented/Unfermented	Genotypes
<ul style="list-style-type: none">• Vis and NIR spectral regions for discrimination• High performances with two pairs of wavelengths (472-636 nm and 2096-2278 nm)• Potential for tailored spectral sensors (smaller, cheaper, handheld) 	<ul style="list-style-type: none">• Great potential of Vis-NIR for discriminating main cocoa genotypes of Pará• Remain cautious : Additional analyses with more samples• Analyses of unfermented beans seem more reliable

————> Vis-NIR = Practical and efficient solution to address challenges related to quality control and authentication

Acknowledgements

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Advances in the individual authentication of cocoa beans: Vis/NIR spectroscopy as a tool to distinguish fermented from unfermented beans and classify genotypes in the eastern Amazonia

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